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DOSE ENHANCEMENT EFFECT OF ANTICANER DRUGS ASSOCIATED WITH INCREASED TEMPERATURE IN VITRO

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Aim: To evaluate in vitro the influence of elevated temperature (42 °C for 60 min) on the action of anticancer drugs doxorubicin, vinorelbine, carboplatin, ifosfamide, etoposide, oxaliplatin, docetaxel and gemcitabine. Methods: HeLa tumor cell cultures, 24 h after seeding, were incubated for 60 min with different concentrations of chemotherapeutical drugs at a temperature of 37 °C or 42 °C. 48 h later the number of viable cells in the flasks were counted using trypan-blue exclusion on a hemacytometer. Results: Hyperthermia alone caused only 10–20% growth inhibition of cell culture. All the chemotherapeutic drugs used demonstrated a dose enhancement effect at elevated temperature. Thermal enhancement ratio for cell proliferation for oxaliplatin, vinorelbine, carboplatin and ifosfamide exceeded 4, for doxorubicin and gemcitabine exceeded 2. Thermal enhancement ratio for cell death did not exceed 1.4. Conclusion: Synergism of hyperthermia and chemotherapeutical drugs was clearly demonstrated for oxaliplatin, vinorelbine, carboplatin, ifosfamide and to a lesser extent for doxorubicin and gemcitabine. Enhancement of the cytostatic effect of anticancer drugs by hyperthermia was more prominent than their cytotoxic effect. Key words: hyperthermia, anticancer drugs, HeLa cells, thermal enhancement ratio.

Multiple studies in hyperthermic oncology testify the enhancing efficacy of combination or multimodality therapy of cancer patients by application of high temperature (40–43 °C for whole-body exposure and 42–47 °C for local treatment). Hyperthermia has been shown to be a powerful modifying factor that increases tumor sensitivity to both radio- and chemotherapy [1–6].

Thermal enhancement of cytotoxic effects for many anticancer drugs has been observed in different experimental systems. The mechanisms involved in the interaction of chemotherapy and hyperthermia are depended on the particular chemotherapeutic agent and thought to be related to an increase in drug uptake, expand of DNA damage and inhibition of DNA repair [1–3, 7]. In vitro drug-induced cell killing usually increased with temperature from 40 °C to 45 °C and maximal cytotoxicity occurred when the drug was scheduled simultaneously with hyperthermia. In vivo the therapeutic efficacy of the thermotherapy was maximized at mild temperature (40.5–43 °C) and the greatest potentiation occurred when the two modalities were administered close together [2, 6, 7].

Despite several experimental studies on the interaction of hyperthermia with chemotherapeutic drugs data concerning treatment efficiency are often conflicting. The use of various cell lines, different treatment schemes and differing end points complicates the interpretation of research findings. Drug concentration and drug exposure time are also of great importance. Furthermore, in vitro and vivo results often differ due to physiology factors of tumor tissue such as microcirculation, pH and oxygenation.

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*Abbreviations used: IC50 (IC90) – the drug concentration causing a 50% (90%) growth inhibition; LC50 – the drug concentration causing a 50% cell killing; TER – thermal enhancement ratio.

Alkylating agents melphalan, cyclophosphamide and ifosfamide were shown to be the most effective drug at moderately elevated temperatures for many types of tumors, effectiveness of cisplatin or oxaliplatin was less pronounced [8, 9, 10]. Hyperthermia was proved to enhance the sensitivity of human tumor cells to doxorubicin [7, 11, 12], vincristine [12] and carboplatin [13, 14], whereas the efficacy of etoposide is only slightly influenced by additional application of heat [15, 16] or decreased [17]. However it was demonstrated that appropriately scheduled hyperthermia and etoposide treatments result in increased etoposide-induced death of human leukemia cells [18]. Simultaneous application of gemcitabine and heat led to decreased cytotoxicity [19, 20], had not the influence on cytotoxicity [12] or augmented cytotoxic effect of drug [10, 21, 22]. There are also contradictory reports as to the interaction of hyperthermia and the taxanes [10, 23-28].

Therefore it is important to compare in equal treatment conditions the impact of hyperthermia on the action of the main chemotherapeutic drugs currently used for cancer treatment. In this study we investigated the differences in HeLa tumor cell culture growth after incubation with different anticancer drugs at a temperature of 37 °C or 42 °C.

MATERIALS AND METHODS

Chemotherapeutic drugs. The drugs investigated were ifosfamide (holoxan, Baxter Oncology Gmbx, Germany), doxorubicin (doxorubicin hydrochloride, RUE "Belmedpreparaty", Belarus), carboplatin (carboplatin-LENS, Veropharm, Russia), oxaliplatin (eloxatin, Sanofi-Synthelabo, France), vinorelbin (neocitec, Labinca, Argentina), etoposide (etoposide-LENS, Veropharm, Russia), docetaxel (docetal, Dabur Pharma Ltd., India), gemcitabine (gemzar, "Eli Lilly", USA). Just before use, the drugs were diluted with culture medium.

Cell culture. HeLa cells (human cervical carcinoma) were obtained from the cell culture collection of Research Institute of Epidemiology and Microbiology (RIEM, Minsk, Belarus). The cells were cultured as a monolayer in culture flasks covered with 199 medium (RIEM) supplemented with 10% fetal calf serum (RIEM) and 100 μg/ml kanamycin.

Cell treatment. One day before the drug treatment, cell monolayer was detached from the culture flasks with 0.02% versen (DIALEK, Belarus) and plated at a concentration of 2×10^5 cells/flask. All the experiments were carried out on exponentially growing cells. On the day of the experiment, $100 \, \mu l$ of each drug dilutions were added into the flasks with cell monolayer and then the cells were incubated in water bath at $37.0\,^{\circ}\text{C} \pm 0.1\,^{\circ}\text{C}$ or $42.0\,^{\circ}\text{C} \pm 0.1\,^{\circ}\text{C}$ for 60 min. Treated and control cells were cultured for additional 48 h, then cell monolayers were versenized and viable cells were counted using trypan-blue exclusion on a haemocytometer. The mean \pm standard deviation was calculated from three experiments.

To differentiate between cytostatic and cytotoxic effects, the number of viable cells after the treatment was compared with the initial cell number before any treatment. Percentage of viable cells above the initial cell number denoted a cytostatic effect, whereas those below showed cytotoxicity [23]. The percentages of viable cells for each treatment were calculated from $100 \times [(N_t - N_{_{\scriptsize o}}) / (N_{_{\scriptsize o}} - N_{_{\scriptsize o}})]$, if N_t was greater than or equal $N_{_{\scriptsize o}}$, or $100 \times [(N_t - N_{_{\scriptsize o}}) / N_{_{\scriptsize o}}]$, if N_t was less than $N_{_{\scriptsize o}}$ (N_t — cell number in treated cultures with chemotherapeutic drug and/or hyperthermia; N_k — cell number in control cultures that had not been exposed to chemotherapeutic drugs or hyperthermia; $N_{_{\scriptsize o}}$ — initial cell number before the treatment).

The values of IC $_{50}$ and IC $_{90}$, concentrations of each drug required for a 50% and 90% reduction in cell number relative to control value, and LC $_{50}$, concentration of drug required for a 50% reduction in cell number relative to initial value before the treatment, were determined for each experiment using regression analysis of the data received. Thermal enhancement ratios for inhibition of cell proliferation and cell killing for each chemotherapeutic drug were calculated as IC $_{50}$, IC $_{90}$, or LC $_{50}$ for drug alone divided by IC $_{50}$, IC $_{90}$, or LC $_{50}$ for drug combined with hyperthermia.

RESULTS AND DISCUSSION

The survivals of exponentially growing HeLa cells exposed to various concentration of ifosfamide doxorubicin, carboplatin, docetaxel, vinorelbine, etoposide, oxaliplatin or gemcitabine at 37 °C or 42 °C are shown in Figure. These data demonstrate that cell number reduced with increasing concentrations of chemotherapeutic agents. All used drugs demonstrated the dosedependent enhancement of cytotoxicity at elevated temperature — the decrease of cell number at 42 °C was more pronounced than at 37 °C. It must be noted the heat treatment alone did not alter significantly cell number causing only 10–20% growth inhibition.

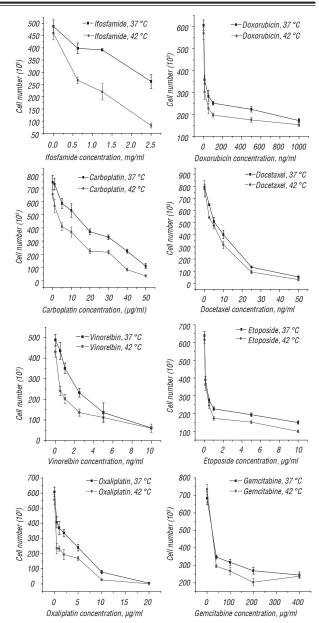


Figure. Dose-response curves for HeLa cells treated with ifosfamide, doxorubicin, carboplatin, oxaliplatin, vinorelbine, etoposide, docetaxel or gemcitabine at $37\,^{\circ}\mathrm{C}$ or $42\,^{\circ}\mathrm{C}$. The cells were incubated with different concentration of anticancer drug for 1 h at $37\,^{\circ}\mathrm{C}$ or $42\,^{\circ}\mathrm{C}$ and counted $48\,\mathrm{h}$ later. Each data point represents the mean of three separate experiments. Bars are standard errors

The parameters of cytostatic (IC_{50} and IC_{90}) and cytotoxic (LC₅₀) effects of chemotherapeutic drugs at 37 °C and 42 °C are presented in Table. The data obtained testify the enhanced efficiency at elevated temperature to a greater or lesser extent of all the drugs under investigation. So, by IC₅₀ and IC₉₀ criteria, hyperthermia caused significant enhancement of cell culture growth inhibition induced by ifosfamide, carboplatin, vinorelbine, oxaliplatin, with thermal enhancement ratio exceeded 4. The values of IC₅₀ and IC₉₀ decreased actually for vinorelbine 5.5-fold and for oxaliplatin 7.2-fold after combination with heat. To a lesser degree hyperthermia enhanced cytostatic effect of doxorubicin and gemcitabine, with thermal enhancement ratio exceeded 2. The differences in ${\rm IC}_{\rm 50}$ and ${\rm IC}_{\rm 90}$ for docetaxel and etoposide were not significant statistically. By LC₅₀ criterion, statistically

Table. The values of IC_{so,1} IC_{so,2} and LC_{so,3} of dose-response curves for HeLa cells treated with different chemotherapeutic drugs at a temperature of 37 °C or 42 °C

Drug	IC50			IC90			LC50		
	37 °C	42 °C	TER	37 °C	42 °C	ER	37 °C	42 °C	TER
Ifosfamide (mg/ml)	1.7 ± 0.1	0.4 ± 0.1*	4.3	> 2.5	1.1 ± 0.1*	> 2.3	> 2.5	2.3 ± 0.1	> 1.1
Doxorubicin (ng/ml)	8.1 ± 1.8	8.1 ± 1.6	1	106.2 ± 1.8	$37.7 \pm 1.6*$	2.8	> 1000	> 1000	_
Carboplatin (µg/ml)	11.7 ± 1.7	$2.7 \pm 1.9*$	4.3	32.1 ± 1.7	14.1 ± 1.9*	2.3	48.9 ± 1.7	$36.8 \pm 1.9*$	1.3
Vinorelbine (ng/ml)	1.1 ± 0.3	$0.2 \pm 0.3^*$	5.5	2.5 ± 0.3	$0.6 \pm 0.3*$	4.2	7.4 ± 0.3	$6.0 \pm 0.3^*$	1.2
Docetaxel (ng/ml)	5.6 ± 1.9	4.2 ± 2.3	1.3	18.0 ± 1.9	13.9 ± 2.3	1.3	35.2 ± 1.9	$24.6 \pm 2.3^*$	1.4
Etoposide (µg/ml)	0.1 ± 0.1	0.1 ± 0.1	1.0	0.7 ± 0.1	0.5 ± 0.1	1.4	> 10	9.2 ± 0.1	> 1.1
Oxaliplatin (µg/ml)	0.4 ± 0.6	0.2 ± 0.2	2.0	3.6 ± 0.6	$0.5 \pm 0.2*$	7.2	8.6 ± 0.6	$6.5 \pm 0.2^*$	1.3
Gemcitabine (µg/ml)	26.8 ± 20.2	25.3 ± 12.8	1.1	187.7 ± 20.2	84.4 ± 12.8*	2.2	> 400	> 400	_

TER – thermal enhancement ratio calculated as IC $_{50}$, IC $_{90}$, or LC $_{50}$ for drug alone divided by IC $_{50}$, IC $_{90}$, or LC $_{50}$ for drug combined with hyperthermia. *The difference between the values for 37 °C and 42 °C are statistically significant, with $\rho < 0.01$.

significant enhancement of drug-mediated cytotoxicity under elevated temperature was observed only for carboplatin, vinorelbine, docetaxel and oxaliplatin, with thermal enhancement ratio for cell death did not exceed 1.4. Thus, thermal enhancement for drug-mediated inhibition of cell proliferation by combination of chemotherapeutic drugs with hyperthermia was more pronounced than for drug-mediated cell killing. The most efficient chemotherapeutic drugs in HeLa cells at elevated temperature 42 °C were ifosfamide, carboplatin, vinorelbine and oxaliplatin.

In this study we investigated in cell culture the action of eight anticancer drugs in broad range of concentrations to describe their both cytostatic and cytotoxic effects. To quantitatively evaluate the impact of the hyperthermia on the chemosensitivity of tumor cells we compared the values of IC_{50} , IC_{90} , and LC_{50} for dose-response curves of HeLa cell culture treated with chemotherapeutic drugs alone or in combination with heat. It was found that interaction of hyperthermia and simultaneous chemotherapy depends on the drug concentration. At low doses of chemotherapeutic agents that exerted mainly cytostatic effect it was shown synergistic interaction with heat for all the drugs under investigation except for docetaxel and etoposide, whereas at high doses of drugs that exerted predominantly cytotoxic action their interaction with heat was only additive.

Thermal enhancement of the cytostatic effect we have observed in this study in vitro may be of importance under in vivo conditions where events inducing the delay of tumor growth are expected to be cytostatic rather than cytotoxic [10]. We believe the data present in this study have clinical implication having recently begun animal tumor experiments to evaluate efficiency of the same chemotherapy in protocols with wholebody hyperthermia.

REFERENCES

- 1. **Zhavrid EA, Osinsky SP, Fradkin SZ.** Hyperthermia and hyperglicemia in oncology. Kiev: Naukova dumka, 1987. 256 p (In Russian).
- 2. **Urano M, Kuroda M, Nishimura Y.** For the clinical application of thermochemotherapy given at mild temperatures. Int J Hyperthermia 1999, **15**: 79–107.
- 3. **Falk MH, Issels RD.** Hyperthermia in oncology. Int J Hyperthermia 2001, **17**: 1–18.
- 4. **Fradkin SZ, Zhavrid EA, Istomin YuP.** Promising trends in the development of systemic hyperthermia in multimodality therapy for malignant neoplasms. In: Book of Abstracts 23-d Annual Meeting of the European Society for Hyperthermic Oncology. Berlin, 2006: 22–4.

- 5. Shevchenko AI, Ganul VL, Ganul AV, Osinsky DS. Hyperthermia in oncology: achievements and original methods. Oncology 2006, **8:** 222–7 (In Ukrainian).
- 6. **Sugarbaker PH.** Laboratory and clinical basis for hyperthermia as a component of intracavitary chemotherapy. Int J Hyperthermia 2007, **23**: 431–42.
- 7. **Hahn GM.** Potential for therapy of drugs and hyperthermia. Cancer Res 1979, **39**: 2264–8.
- 8. **Urano M, Ling CC.** Thermal enhancement of melphalan and oxaliplatin cytotoxicity in vitro. Int J Hyperthermia 2002, **18**: 307–15.
- 9. Takemoto M, Kuroda M, Urano M, Nishimura Y, Kawasaki S, Kato H, Okumura Y, Akaki S, Kanazawa S, Asaumi J, Joja I, Hiraki Y. The effect of various chemotherapeutic agents given with mild hyperthermia on different types of tumours. Int J Hyperthermia 2003, 19: 193–203.
- 10. Mohamed F, Marchettini P, Stuart OA, Urano M, Sugarbaker PH. Thermal enhancement of new chemotherapeutic agents at moderate hyperthermia. Ann Surg Oncology 2003, 10: 463–8.
- 11. **Urano M, Begley J, Reynolds R.** Interaction between adriamycin cytotoxicity and hyperthermia: growth-phase-dependent thermal sensitization. Int J Hyperthermia 1994, **10**: 817–26.
- 12. **Hermisson M, Weller M.** Hyperthermia enhanced chemosensitivity of human malignant glioma cells. Anticancer Res 2000, **20**: 1819–23.
- 13. Choi EK, Park SR, Lee JH Chung HS, Ahn HE, Rhee YH, Lim BU, Park HJ. Induction of apoptosis by carboplatin and hyperthermia alone or combined in WERI human retinoblastoma cells. Int J Hyperthermia 2003, **19**: 431–43.
- 14. **Schem BC**, **Mella O**, **Dahl O**. Thermochemotherapy with cisplatin or carboplatin in the BT4 rat glioma in vitro and in vivo. Int J Radiat Oncol Biol Phys 1992, **23**: 109–14.
- 15. **Debes A, Willers R, Gobel U, Wessalowski R.** Role of heat treatment in childhood cancers: distinct resistance profiles of solid tumor cell lines towards combined thermochemotherapy. Pediatr Blood Cancer 2005, **45**: 663–9.
- 16. **Souslova T, Averill-Bates DA.** Multidrug-resistant hela cells overexpressing MRP1 exhibit sensitivity to cell killing by hyperthermia: Interactions with etoposide. Int J Rad Onc Biol Phys 2004, **60**: 1538–51.
- 17. **Dynlacht JR, Wong RS, Albright N, Dewey WC.** Hyperthermia can reduce cytotoxicity from etoposide without a corresponding reduction in the number of topoisomerase II-DNA cleavable complexes. Cancer Res 1994, **54**: 4129–37.
- 18. **Pantazis P, Han Z, Wyche J.** Schedule-dependent efficiency of thermochemotherapy in vitro with etoposide and heating at 43 degrees C. Anticancer Res 1999, **19**: 995–8.
- 19. Haveman J, Rietbroek RC, Geerdink A, Van Rijn J, Bakker PJ. Effect of hyperthermia on the cytotoxicity of 2',2'-difluorodeoxycytidine (gemcitabine) in cultured SW1573 cells. Int J Cancer 1995, **62**: 627–30.
- 20. Van Bree C, Beumer C, Rodermond HM, Haveman J, Bakker PJ. Effectiveness of 2',2'difluorodeoxycytidine

(Gemcitabine) combined with hyperthermia in rat R-1 rhabdomyosarcoma in vitro and in vivo. Int J Hyperthermia 1999, **15**: 549–56.

- 21. **Van der Heijden AG, Verhaegh G, Jansen CF, Schalken JA, Witjes JA.** Effect of hyperthermia on the cytotoxicity of 4 chemotherapeutic agents currently used for the treatment of transitional cell carcinoma of the bladder: an in vitro study. J Urol 2005, **173**: 1375–80.
- 22. Vertrees RA, Das GC, Popov VL, Coscio AM, Goodwin TJ, Logrono R, Zwischenberger JB, Boor PJ. Synergistic interaction of hyperthermia and Gemcitabine in lung cancer. Cancer Biol Ther 2005, **10**: 1144–53.
- 23. Michalakis J, Georgatos SD, Romanos J, Koutala H, Georgoulias V, Tsiftsis D, Theodoropoulos PA. Micromolar taxol, with or without hyperthermia, induces mitotic catastrophe and cell necrosis in HeLa cells. Cancer Chemother Pharmacol 2005, **56**: 615–22.
- 24. Michalakis J, Georgatos SD, de Bree E, Polioudaki H, Romanos J, Georgoulias V, Tsiftsis DD, Theodoropoulos PA.

- Short-term exposure of cancer cells to micromolar doses of paclitaxel, with or without hyperthermia, induces long-term inhibition of cell proliferation and cell death in vitro. Ann Surg Oncol 2007, **14**: 1220–8.
- 25. Othman T, Goto S, Lee JB, Taimura A, Matsumoto T, Kosaka M. Hyperthermic enhancement of the apoptotic and antiproliferative activities of paclitaxel. Pharmacology 2001, **62**: 208–12.
- 26. Rietbroek RC, Katschinski DM, Reijers MH, Robins HI, Geerdink A, Tutsch K, d'Oleire F, Haveman J. Lack of thermal enhancement for taxanes in vitro. Int J Hyperthermia 1997, 13: 525–33.
- 27. **Leal BZ, Meltz ML, Mohan N, Kuhn J, Prihoda TJ, Herman TS.** Interaction of hyperthermia with Taxol in human MCF-7 breast adenocarcinoma cells. Int J Hyperthermia 1999, **15**: 225–36.
- 28. **Mohamed F, Stuart OA, Glehen O**, *et al.* Docetaxel and hyperthermia: factors that modify thermal enhancement. J Surg Oncol 2004, **88**: 14–20.

ЭФФЕКТ УСИЛЕНИЯ АКТИВНОСТИ ПРОТИВООПУХОЛЕВЫХ ПРЕПАРАТОВ ПРИ ПОВЫШЕНИИ ТЕМПЕРАТУРЫ *IN VITRO*

Цель: изучить *in vitro* влияние повышенной температуры (42 °C в течение 60 мин) на действие противоопухолевых препаратов: доксорубицина, винорельбина, карбоплатина, ифосфамида, этопозида, оксалиплатина, доцетаксела и гемцитабина. *Методы:* культуру опухолевых клеток HeLa через 24 ч после рассева инкубировали в течение 60 мин с различными концентрациями химиотерапевтических препаратов при температуре 37 °C или 42 °C. Спустя 48 ч подсчитывали количество живых клеток во флаконах, используя гемоцитометр и метод исключения красителя трипанового синего. *Результаты:* гипертермия сама по себе вызывала 10−20% угнетение роста культуры клеток. У всех исследованных химиотерапевтических препаратов отмечали эффект усиления при повышенной температуре. Коэффициент теплового усиления в отношении клеточной пролиферации для оксалиплатина, винорельбина, карбоплатина и ифосфамида превысил 4,0, для доксорубицина и гемцитабина — 2,0. Коэффициент теплового усиления в отношении гибели клеток не превышал 1,4. *Выводы:* синергизм гипертермии и химиотерапевтических препаратов продемонстрирован для оксалиплатина, винорельбина, карбоплатина, ифосфамида, в меньшей степени — для доксорубицина и гемцитабина. Усиление цитостатического эффекта противоопухолевых препаратов под действием гипертермии было более выраженным, чем их цитотоксического эффекта.

Ключевые слова: гипертермия, противоопухолевые препараты, клетки HeLa, коэффициент теплового усиления.