

ROLE OF VASCULAR ENDOTHELIAL GROWTH FACTOR IN NON-SMALL CELL LUNG CANCER PATHOGENESIS

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The angiogenesis is an important process in the pathogenesis of malignancies. It is regulated by various growth factors, with the vascular endothelial growth factor (VEGF) playing the central role. *The aim* of the present study was to evaluate possible associations of functional *VEGF* –2578C>A, –634G>C, and +936C>T polymorphisms with the risk for occurrence and progression of non-small cell lung cancer (NSCLC) in patients living in Republic of Belarus. *Materials and Methods:* A total of 202 patients (147 males and 55 females) diagnosed as having the NSCLC. The control group consisted of 336 individuals (245 males and 91 females) without an oncopathology. The total DNA was isolated from peripheral blood. We investigated the single nucleotide polymorphisms of *VEGF* (rs 2010963), (rs 699947), (rs 3025039). The genotyping was performed by PCR-RFLP analysis. *Results:* Our results revealed a marginally significant association of the –2578CC genotype ($p=0.002$) with a greater degree of tumor spread (T2–T4). Heterozygous genotypes –2578CA and +936CT carriers were included into the follow-up group significantly more often ($p=0.021$ and $p=0.012$, respectively). Our study demonstrate that *VEGF* –2578A/C and +936C/T polymorphisms are among the factors determining the individual peculiarities of NSCLC course in this population and can be used for clarifying the prognosis of the disease.

Key Words: vascular endothelial growth factor, polymorphism, non-small cell lung cancer.

The angiogenesis is an important process in the pathogenesis of malignancies. It is regulated by various growth factors, with the vascular endothelial growth factor (VEGF-A or VEGF) playing the central role [1]. The VEGF gene expression is closely related to the degree of vascularization and the prognosis for the occurrence of numerous solid tumors and is considered to be a predictor of resistance to the chemo- and radiotherapy [2]. An elevated VEGF expression is associated with tumor growth and metastatic process, while the inhibited VEGF expression results in suppressed tumor growth [3].

The *VEGF* gene also triggers the activation of the protease cascade involved in the degradation of extracellular matrix, suppressing apoptosis, stimulates the endothelial cells survival, increases vascular permeability, inhibits the dendritic cells differentiation, regulates the hexose transport in endothelial cells as well as activates tissue factors and monocytes migration [1]. Clinical studies have shown that the high level of VEGF expression and, respectively, the increased number of microvessels in the tumor correlate with the disease stage and the unfavorable prognosis for many tumor types, including the non-small cell lung cancer (NSCLC) [4–8].

The *VEGF* gene is located on the short arm of chromosome 6 (6p21.3) and consists of eight exons and seven introns [7]. VEGF is a diametric glycoprotein, acting via tyrosine kinase receptors

VEGFR1 and VEGFR2 located predominantly on endothelial cells [9].

Polymorphic sites of *VEGF* gene determining the level of VEGF production are found in the promoter — 5'-untranslated region (5'-UTR) as well as in 5'- and 3'-untranslated regions of the gene (3'-UTR).

The VEGF gene polymorphism is associated with differentiated VEGF expression and protein production. *VEGF*-2578C/C, –634 C/-genotypes are related to high level of VEGF expression [5, 9–12], while +936T allele correlates with low VEGF expression and its low levels in blood plasma [13, 14].

Recent studies show that individual polymorphic variants of *VEGF* influence the development of certain cancers. In particular, *VEGF* C-634G, C-2578A, C+936T polymorphisms are associated with an increased lung cancer risk in Asian population [15]. Based on the above-mentioned, it can be assumed that these single nucleotide substitutions of the *VEGF* gene related to angiogenesis can affect the risk of tumour occurrence and progression as well as the patients' survival.

The aim of the present study was to evaluate possible associations of functional *VEGF* –2578C>A, –634G>C, and +936C>T polymorphisms with the risk for occurrence and progression of NSCLC in patients living in the territory of the Republic of Belarus.

MATERIALS AND METHODS

202 patients (147 males and 55 females) diagnosed as having the NSCLC and treated at the Minsk City Oncology Dispensary during the period from 2003 to 2012 were included in the study. The control group consisted of 336 individuals (245 males and 91 females) without an oncopathology who were age-, gender- and comorbidity matched with NSCLC pa-

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Abbreviations used: 3'-UTR – 3'-untranslated region; 5'-UTR – 5'-untranslated region; NSCLC – non-small cell lung cancer; VEGF – vascular endothelial growth factor.

tients. Clinical characteristics of patients with NSCLC and controls are presented in Table 1.

Table 1. Characteristics of study population

Characteristics	Patients with NSCLC	Controls
	(n = 202) n (%)	(n = 336) n (%)
Gender:		
female	55 (27.2)	91 (27.1)
male	147 (72.8)	245 (72.9)
Smoking status:		
smokes	67 (33.2)	201 (59.8)
does not smoke	119 (58.9)	135 (40.2)
no information	16 (7.9)	–
Stage:		
I	108 (53.5)	–
II	27 (13.4)	–
III	55 (27.2)	–
IV	12 (5.9)	–
Histology:		
squamous-cell carcinoma	106 (52.5)	–
adenocarcinoma	96 (47.5)	–
Surgery:		
lobectomy/bilobectomy	121 (65.1)	–
pneumonectomy	42 (22.6)	–
lung resection	13 (7.0)	–
biopsy	10 (5.4)	–
Therapy:		
chemotherapy	65 (32.2)	–
radiotherapy	39 (19.3)	–
no therapy	98 (48.5)	–

The study was performed in compliance with the principles of voluntary participation and confidentiality, in accordance with the questioning of patients and the approval from the local Ethics Committee to study tissue samples and biological fluids.

The diagnosis of lung cancer in patients has been established on the basis of clinical signs of the disease, history data, bronchoscopy, X-ray examination and computed tomography, cytomorphology of sputum and tumour tissue biopsies. In the group of NSCLC patients, the mean age was 61.6±0.6, in the control group — 61.6±0.8 years.

All cases of lung malignancies have been identified according to the International Classification TNM/pTNM (7th edition, 2009). The histological type of lung carcinoma was determined according to the WHO histological criteria (3rd edition, 1999). The group of NSCLC included the most common neoplasms — squamous-cell carcinoma and adenocarcinoma. The treatment was mostly surgical: lobectomy — 65.1%, pneumonectomy — 22.6%,

lung resection — 7.0%. Only a biopsy was performed in 5.4% of cases, and in 7.9% of patients the operation was not done because of the process spread or severe comorbidity.

The total DNA from the peripheral blood was isolated using the Mathew method [16]. *VEGF* C-634G (rs 2010963), C-2578A (rs 699947), C+936T (rs 3025039) polymorphisms were genotyped by the polymerase chain reaction method and the restriction fragment length polymorphism analysis (PCR-RFLP analysis) using specific primers and restriction endonucleases [15, 17]. Primers for the analysis were synthesized by “Primetekh”, Minsk. The reagents for PCR and PCR-RFLP were manufactured by “Fermentas”, Vilnius.

Statistical analysis. Statistical analyses were performed using Excel and Statistica 7.0. When comparing genotype frequencies, Pearson’s chi-squared test (χ^2) was used. The association between the genotypes and the disease course was assessed by the value of odds ratio (OR).

RESULTS AND DISCUSSION

An elevated VEGF expression was revealed while studying the variety of malignancies: cancers of the colon, rectum, liver, lung, thyroid, digestive tract; breast, kidney and bladder adenocarcinoma; ovary and uterus carcinoma; angiosarcoma, multi-form glioblastoma [18–21]. In turn, the level of VEGF expression depends largely on the polymorphic variants of the gene.

Therefore, the first phase of our work was to analyse the association between *VEGF* C-634G, C-2578A and C+936T polymorphisms and the risk of NSCLC.

The analysis of data on *VEGF* genotyping in patients with NSCLC and controls is presented in Table 2.

Frequency distribution of alleles and genotypes of functionally significant *VEGF* polymorphisms in the population of patients living in the territory of the Republic of Belarus was close to the occurrence of the same polymorphisms in the control group. Statistically significant differences in the frequency of the studied genotypes between the group of patients with NSCLC and the control group were not

39. Schultz A, Lavie L, Hochberg I, *et al.* Interindividual heterogeneity in the hypoxic regulation of VEGF: significance for the development of the coronary artery collateral circulation. *Circulation* 1999; **100**: 547–52.

found. Similar results on the frequency of the studied *VEGF* single nucleotide substitutions were obtained for the populations in Germany and Sweden.

Based on the results, it can be assumed that there is no connection between the studied *VEGF* polymorphisms and the risk of NSCLC in this population.

Since *VEGF* is a major mediator of angiogenesis, it can be supposed that *VEGF* C-634G, C-2578A and C+936T polymorphisms will rather affect the phenotype and tumor biological behavior than the risk for tumor occurrence.

The next stage of our study was to find the relationship between the polymorphisms under study and the clinical course of this disease. The comparative analysis of genotypes distribution of three functionally significant *VEGF* polymorphisms and tumor size, metastasis and clinical outcomes in the group of patients with NSCLC has been carried out.

VEGF triggers the neoplastic angiogenesis, resulting in increased microvascular density, and malignant tissue receives more nutrients [23, 24]. *VEGF* secretion by tumor cells leads to the synthesis of proangiogenic factors. The newly formed vessels begin to supply malignant tissue with oxygen and nutrients; the tumor is growing and producing more *VEGF*. *VEGF* increases the level of *VEGFR2* receptor expression by endothelial cells of tumor microvessels, thus stimulating the cell growth and endothelial cells proliferation [25].

Structurally and functionally the neoplastic vessels differ from the normal ones, with high permeability, chaotic branching, multiple loops, weaves, dead-end branches, lack of structured vasculature being typical for them [2]. Chaotic arrangement of tumor vessels results in uneven oxygen supply to the surrounding tissues, formation of local hypoxic foci, and is followed by tumor resistance to the radio- and chemotherapy.

With an elevated *VEGF* expression, the vascular permeability increases leading to a higher interstitial

and intratumoural pressure, facilitating the penetration of tumor cells into the bloodstream [26].

The relationship between the tumor angiogenesis level, tumor size and metastatic process is confirmed by the correlation between the course of the disease and microvascular density of the primary tumor [27–32]. Different *VEGF* polymorphisms are associated with lymphogenic and hematogenous metastases in a variety of malignancies [28, 29].

The performed analysis of the relationships between three *VEGF* gene polymorphisms and tumor size (Table 3) has shown that genotype –2578CC carriers displayed a greater degree of tumor spread (T2–4) significantly more frequently ($p = 0,002$) compared to the primary focus spread (T1). On the contrary, the carriers of –2578CA genotype more often presented with small non-invasive cancer (T1). However, in this group of NSCLC patients, no statistically significant associations between the studied polymorphic allele variants and regional and/or distant metastases were found (Table 4).

The *in vitro* studies conducted by M. Mohammadi and M.C. Shahbazi has shown that -2578C allele correlated with higher *VEGF* expression compared to A allele [30, 31]. Similar results were obtained by M. Perrot-Applanat who proved that –2578C allele was associated with an elevated *VEGF* expression and pointed to the possible involvement of *VEGF* proteins in the autocrine regulation of the tumor growth [32]. Perhaps, this is likely to explain our findings, where the homozygous carriers (–2578CC) had higher degree of the tumor spread. The lower level of *VEGF* expression in the presence of –2578A allele seems to have a protective effect in heterozygous carriers (–2578CA) associated with a lesser degree of the neoplasm spread.

In the population under study, a significant relationship between the –2578CA variant and the degree of spread of the primary tumor as well as the outcome of the disease was detected (Table 5). Heterozy-

Table 2. Frequency distribution of polymorphic variants of *VEGF* in patients with NSCLC and control groups in different populations

Genotype	Belarus (original data)			Germany [22]			Sweden [22]		
	Patients N=186; n (%)	Controls N=364; n (%)	OR (95% CI)	Patients N, %	Controls N, %	OR (95% CI)	Patients N=936; n (%)	Controls N=941; n (%)	OR (95% CI)
<i>VEGF</i> (G634C)									
GG	83 (44.6)	186 (51.1)	0.77 (0.54–1.10)	–	–	–	488 (52.1)	492 (52.3)	1.00
GC	88 (47.3)	154 (42.3)	1.22 (0.86–1.75)	–	–	–	363 (38.8)	367 (39.0)	1.00 (0.82–1.21)
CC	15 (8.1)	24 (6.6)	1.24 (0.64–2.43)	–	–	–	85 (9.1)	82 (8.7)	1.05 (0.74–1.47)
G allele	254 (68.3)	526 (72.3)	0.83 (0.63–1.09)	–	–	–	1339 (71.5)	1351 (72.0)	0.99 (0.86–1.14)
C allele	118 (31.8)	202 (27.7)	1.21 (0.92–1.59)	–	–	–	533 (28.5)	531 (28.0)	1.01 (0.86–1.17)
<i>VEGF</i> (C2578A)									
CC	41 (25.3)	83 (23.0)	1.13 (0.74–1.74)	44 (28.8)	50 (30.9)	1.00	258 (27.5)	257 (27.3)	1.00
CA	90 (55.6)	186 (51.7)	1.17 (0.81–1.70)	75 (49.0)	72 (44.4)	1.18 (0.68–2.06)	449 (47.8)	451 (48.0)	0.99 (0.79–1.24)
AA	31 (19.1)	91 (25.3)	0.70 (0.44–1.11)	34 (22.2)	40 (24.7)	0.97 (0.50–1.86)	232 (24.7)	232 (24.7)	1.00 (0.82–1.23)
C allele	172 (53.1)	352 (48.9)	1.18 (0.91–1.54)	163 (53.3)	172 (53.1)	1.01 (0.74–1.38)	965 (51.4)	965 (51.2)	0.78 (0.68–0.89)
A allele	152 (46.9)	368 (51.1)	0.85 (0.65–1.10)	143 (46.7)	152 (46.9)	0.99 (0.73–1.36)	913 (48.6)	915 (48.8)	1.05 (0.93–1.20)
<i>VEGF</i> (C936T)									
CC	115 (71.4)	267 (74.2)	0.87 (0.57–1.32)	120 (78.4)	128 (78.5)	1.00	708 (76.6)	720 (77.1)	1.00
CT	38 (23.6)	80 (22.2)	1.08 (0.70–1.68)	31 (20.3)	31 (19.0)	1.07 (0.59–0.93)	204 (22.1)	203 (21.7)	1.02 (0.81–1.28)
TT	8 (5.0)	13 (3.6)	1.40 (0.57–3.44)	2 (1.3)	4 (2.5)	0.53 (0.07–3.46)	12 (1.3)	11 (1.2)	1.11 (0.45–2.71)
C allele	268 (83.2)	614 (85.3)	0.86 (0.60–1.23)	271 (59.8)	287 (88.0)	0.20 (0.14–0.30)	1620 (87.7)	1643 (88.0)	0.97 (0.80–1.18)
T allele	54 (16.8)	106 (14.7)	1.17 (0.82–1.67)	182 (40.2)	39 (12.0)	4.94 (3.37–7.25)	228 (12.3)	225 (12.0)	1.03 (0.84–1.25)

Table 3. Distribution of frequencies of polymorphic variants of *VEGF* and tumor size in patients with NSCLC

Tumour size	N	G634C						N	C2578A						N	C936T					
		GG		GC		CC			CC		CA		AA			CC		CT		TT	
		n	%	n	%	n	%		n	%	n	%	n	%		n	%	n	%	n	%
T1	70	32	45.7	33	47.1	5	7.1	57	6	10.5*	40	70.2**	11	19.3	41	29	70.7	11	26.8	1	2.4

gous genotype –2578CA carriers were included into the follow-up group significantly more often ($p = 0.021$). It should be noted that a similar results was observed for another *VEGF* polymorphism — the heterozygous variant +936CT located in the 3′-UTR ($p = 0.012$). Moreover, C. Oliveira et al. have shown that heterozygous +936CT genotype was increasingly associated with a non-recurrent disease [11]. No associations between the *VEGF* –634G>C polymorphism and the survival of patients with NSCLC have been found.

It has been demonstrated that the polymorphism of a single nucleotide located in the promoter region or 3′- and 5′-UTR affects the protein expression at the transcriptional level [33]. Initially, the hypoxia induced factor (HIF-1) joins *VEGF* gene in the promoter region (–2578CA polymorphism) and increases its expression. Gene polymorphism in this region can weaken or strengthen this interaction, and, therefore, alter the *VEGF* expression (–2578CA) [34] that, in turn, will affect the formation and permeability of microvessels and, finally, the tumor size, metastasis and survival.

Based on the results of the study on the correlation between the polymorphisms and development of the disease in this population, it has been established that the carriers of –2578CC genotype displayed a greater degree of tumor spread (T2–4) significantly more often ($p = 0.002$) while the carriers of –2578CA genotype presented with small non-invasive cancers (T1) more frequently ($p = 0.021$). The carriers of –2578C/A genotype were significantly often observed ($p = 0.021$) in the “follow-up” group. Given that the high level of *VEGF* expression is related to an increased risk of a recurrent disease and shorter survival of subjects with different cancers [35], one may suggest the *VEGF* (–2578C/-) genotype to contribute to the high expression of the corresponding protein product in the development of NSCLC.

The mechanism of most associations between single nucleotide substitutions and clinical behavior of tumors is still largely unclear. It should be taken into account that 5′- and 3′-UTR contain key regulatory elements sensitive to hypoxia [36–38] and contribute to high variability of *VEGF* production [39]. For instance, *VEGF*-634G>C and –2578A>C polymorphisms in the 5′-UTR affect the efficiency of protein translation [33], and 936C>T polymorphism in the 3′-UTR influences the *VEGF* circulating plasma concentrations [13] and tumor tissue expression of *VEGF* [5]. This can explain the protective effect of the *VEGF* +936T allele in breast cancer metastasis (due to the reduced *VEGF* expression) [4, 11].

Thus, the results of our study demonstrate that *VEGF* –2578A/C and +936C/T polymorphisms are among the factors determining the individual peculiarities of NSCLC course in this population and can be used for clarifying the prognosis of the disease.

REFERENCES

- Ferrara N, Gerber HP, LeCouter J. The biology of *VEGF* and its receptors. *Nat Med* 2003; **9**: 669–76.
- Ferrara N. Vascular endothelial growth factors as a target for anticancer therapy. *Oncologist* 2004; **9**: 2–10.
- Ferrara N. *VEGF* and the quest for tumour angiogenesis factors. *Nat Rev Cancer* 2002; **2**: 795–803.
- Jin Q, Hemminki K, Enquist K, *et al.* Vascular endothelial growth factor polymorphisms in relation to breast cancer development and prognosis. *Clin Cancer Res* 2005; **11**: 3647–53.
- Koukourakis MI, Papazoglou D, Giatromanolaki A, *et al.* *VEGF* gene sequence variation defines *VEGF* gene expression status and angiogenic activity in non-small cell lung cancer. *Lung Cancer* 2004; **46**: 293–8.
- Liu D-H, Zhang X-Y, Fan D-M, *et al.* Expression of vascular endothelial growth factor and its role in oncogenesis of human gastric carcinoma. *World J Gastroenterol* 2001; **7**: 500–5.
- Yamamori M, Sakaeda T, Nakamura T, *et al.* Association of *VEGF* genotype with mRNA level in colorectal adenocarcinomas. *Biochem Biophys Res Commun* 2004; **325**: 144–50.
- Lu H, Shu Z-O, Cui Y, *et al.* Association of genetic polymorphisms in the *VEGF* gene with breast cancer survival. *Cancer Res* 2005; **65**: 5015–9.
- Yang B, Cross DF, Ollerenshaw M, *et al.* Polymorphisms of the vascular endothelial growth factor and susceptibility to diabetic microvascular complications in patients with type 1 diabetes mellitus. *J Diabetes Complications* 2003; **17**: 1–6.
- Langsenlehner U, Wolf G, Langsenlehner T, *et al.* Genetic polymorphisms in the vascular endothelial growth factor gene and breast cancer risk. The Austrian «tumour of breast tissue: incidence, genetics, and environmental risk factors» study. *Breast Cancer Res Treat* 2008; **109**: 297–304.
- Oliveira C, Lourenço G, Silva P, *et al.* Polymorphisms in the 5′- and 3′-untranslated region of the *VEGF* gene and sporadic breast cancer risk and clinicopathologic characteristics. *Tumour Biol* 2011; **32**: 295–300.
- Qi M, Huang X, Zhou L, Zhang J. Four polymorphisms of *VEGF* (+405C>G, –460T>C, –2578C>A, and –1154G>A) in susceptibility to psoriasis: a meta-analysis. *DNA and Cell Biology* 2014; **33**: 234–44.
- Renner W, Kotschan S, Hoffmann C, *et al.* A common 936 C/T mutation in the gene for vascular endothelial growth factor is associated with vascular endothelial growth factor plasma levels. *J Vasc Res* 2000; **37**: 443–8.
- Schneider B, Radovich M, Sledge G, *et al.* Association of polymorphisms of angiogenesis genes with breast cancer. *Breast Cancer Res Treat* 2008; **111**: 157–63.
- Li Y, Liang J, Liu X, *et al.* Correlation of polymorphisms of the vascular endothelial growth factor gene and the risk of lung cancer in an ethnic Han group of North China. *Exp Ther Med* 2012; **3**: 673–6.
- Mathew CC. The isolation of high molecular weight eucaryotic DNA. In: *Methods in Molecular Biology*. JMNJ Walker ed. Clifton: Human Press, 1984; **2**: 31–4.
- Lee SJ, Lee SV, Jeon HS, *et al.* Vascular endothelial growth factor gene polymorphisms and risk of primary lung cancer. *Cancer Epidemiol Biomarkers Prev* 2005; **14**: 571–5.
- Lee TH, Avraham HK, Jiang S, *et al.* Vascular endothelial growth factor modulates the transendothelial migration of MDA-MB-231 breast cancer cells through regulation of brain microvascular endothelial cell permeability. *J Biol Chem* 2003; **278**: 5277–84.
- Ferrante M, Pierik M, Henckaerts L, *et al.* The role of vascular endothelial growth factor (*VEGF*) in inflammatory bowel disease. *Inflamm Bowel Dis* 2006; **12**: 870–8.
- Poon RT, Fan ST, Wong J. Clinical implications of circulating angiogenic factors in cancer patients. *J Clin Oncol* 2001; **19**: 1207–25.

21. Rosen LS. Clinical experience with angiogenesis signaling inhibitors: focus on vascular endothelial growth factor (VEGF) blockers. *Cancer Control* 2002; **9**: 36–44.
22. Jin Q, Hemminki K, Enquist K, *et al.* Vascular endothelial growth factor polymorphisms in relation to breast cancer development and prognosis. *Clin Cancer Res* 2005; **11**: 3647–53.
23. Carmeliet P, Jain R. Angiogenesis in cancer and other diseases. *Nature* 2000; **407**: 249–57.
24. Jain RK. Normalization of tumour vasculature: an emerging concept in antiangiogenic therapy. *Science* 2005; **307**: 58–62.
25. Millauer B, Shawver LK, Plate KH, *et al.* Glioblastoma growth inhibited *in vivo* by a dominant-negative Flk-1 mutant. *Nature* 1994; **367**: 576–9.
26. Kopnin BP. Basic properties of the neoplastic cells and the underlying mechanisms of their occurrence. *Pract Oncol* 2002; **3**: 229–35 (in Russian).
27. Folkman J. Angiogenesis. *Annu Rev Med* 2006; **57**: 1–18.
28. Ishigami SI, Arii S, Furutani M, *et al.* Predictive value of vascular endothelial growth factor (VEGF) in metastasis and prognosis of human colorectal cancer. *Br J Cancer* 1998; **78**: 1379–84.
29. Kawakami M, Furuhashi T, Kimura Y, *et al.* Expression analysis of vascular endothelial growth factors and their relationships to lymph node metastasis in human colorectal cancer. *J Exp Clin Cancer Res* 2003; **22**: 229–37.
30. Mohammadi M, Ollier WE, Hutchinson IV. A functional association study of VEGF gene promoter polymorphisms with VEGF expression by stimulated pbm cells. *Hum Immunol* 2003; **64**: 125.
31. Shahbazi M, Fryer AA, Pravica V, *et al.* Vascular endothelial growth factor gene polymorphisms are associated with acute renal allograft rejection. *J Am Soc Nephrol* 2002; **13**: 260–4.
32. Perrot-Applanat M, Di Benedetto M. Autocrine functions of VEGF in breast tumour cells: adhesion, survival, migration and invasion. *Cell Adh Migr* 2012; **6**: 547–53.
33. Schneider BP, Radovich M, Miller KD. The role of vascular endothelial growth factor genetic variability in cancer. *Clin Cancer Res* 2009; **15**: 5297–301.
34. Sheng Z, Guo-Ping W, Cong L, Muxiang Z. Eukaryotic initiation factor 4E (eIF4E) and angiogenesis: prognostic markers for breast cancer. *BMC Cancer* 2006; **6**: 231–43.
35. Awata T, Inoue K, Kurihara S, *et al.* A common polymorphism in the 50-untranslated region of the VEGF gene is associated with diabetic retinopathy in type 2 diabetes. *Diabetes* 2002; **51**: 1635–9.
36. Watson C, Webb N, Bottomley M, Brenchley P. Identification of polymorphisms within the vascular endothelial growth factor (VEGF) gene: correlation with variation in VEGF protein production. *Cytokine* 2000; **12**: 1232–5.
37. Brown LF, Detmar M, Claffey K, *et al.* Vascular permeability factor/vascular endothelial growth factor: A multifunctional angiogenic cytokine. In: *Regulation of Angiogenesis. ID Goldberg, EM Rosen*, eds. Birkhäuser Basel, 1997; **79**: 233–69.
38. Minchenko A, Salceda S, Bauer T, Caro J. Hypoxia regulatory elements of the human vascular endothelial growth factor gene. *Cell Mol Biol Res* 1994; **40**: 35–9.